



# SolNanoTox

**How the solubility of nanoparticles impacts on their behaviour and their potential toxicity?**

**Valérie FESSARD**



# Program between France and Germany



**Up to 42 months; from March 2014 up to August 2017**

**Behaviour, toxicity and mechanism of action of NMs through oral exposure not well established**



**Due to the large panel of NMs, is solubility a key factor in hazard assessment?**



**Study interactions and toxic effects with 2 types of NMs, one soluble and one insoluble**

# ***Choice of MNMs***

**2 types of NMs but of similar size**

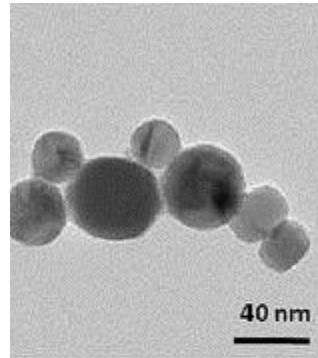
**commonly used in food, or expected to migrate through food processing or materials in contact**



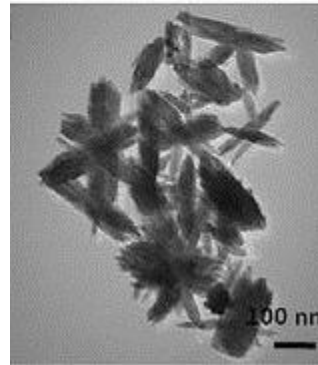
**Will or may be ingested by consumers**

# Choice of MNM

TiO<sub>2</sub>



Anatase



Rutile

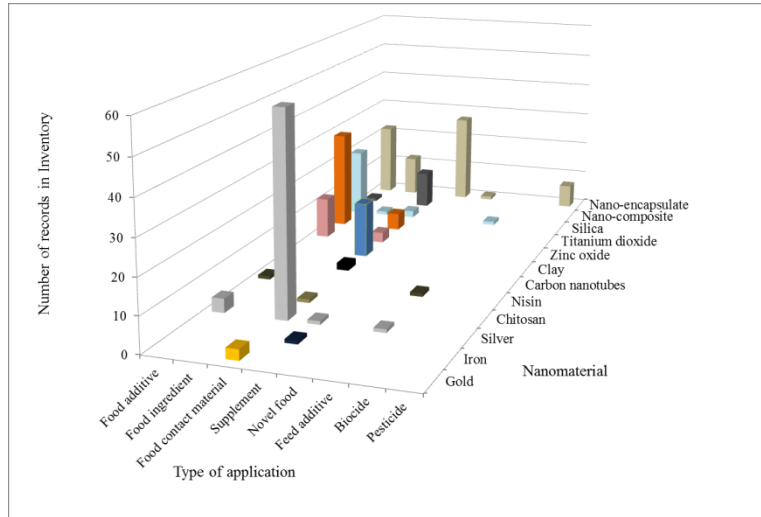
Anatase would produce more oxidative stress and suggested to be more toxic (Xue et al., 2010; Petkovic et al, 2011, review of Wang and Fan, 2014)

But other studies showed that rutile induces higher toxicity (Numano et al , 2014 ; Sund et al, 2014)

Only few data published on the toxicity (especially *in vivo*) of the rutile forms

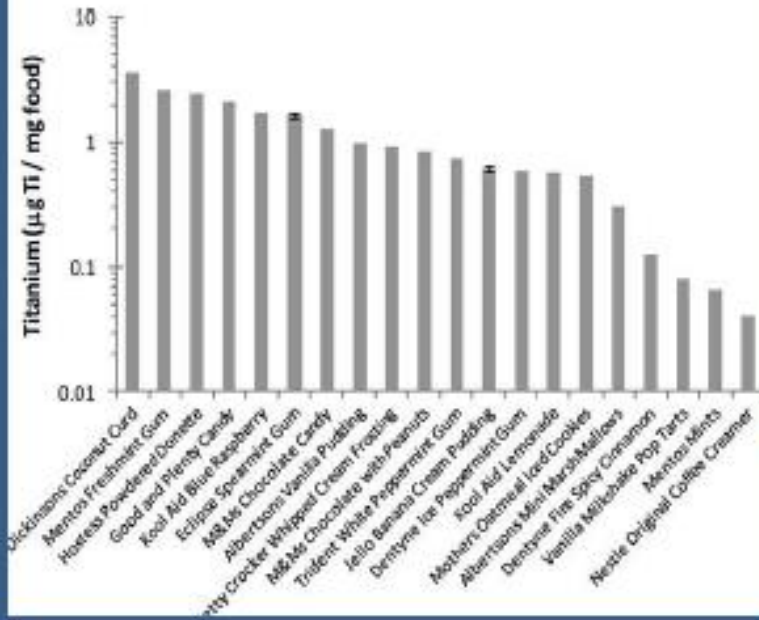
# Choice of MNMs

## TiO<sub>2</sub> uses in food products



NPs applications in the agricultural, feed and food sector  
EFSA report (2014)

Oral  
(0.01 to 2  $\mu\text{g TiO}_2/\text{mg food}$ )



# Choice of MNMs

## TiO<sub>2</sub>: hazardous?

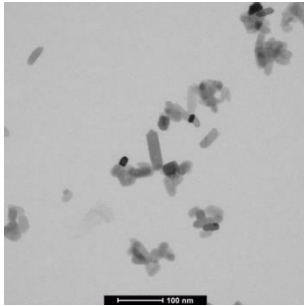
International Agency for Research on Cancer:  
“**possibly carcinogenic to humans**” (2B)

Genotoxicity data *in vitro* and *in vivo*: contradictory results  
Probably oxidative stress induction of DNA damage but not excluded that  
an direct effect may also occur

Biodistribution: some accumulation in liver (Shukla et al 2013; Geraets et al 2014)

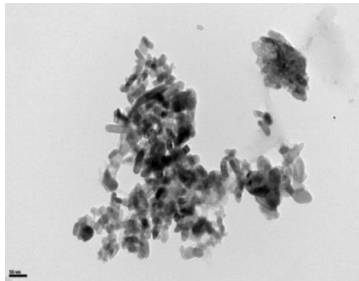
# Choice of MNMs

## TiO<sub>2</sub> (from JRC)



NM103  
(thermal, hydrophobic, coated)

25 nm



NM104  
(thermal, hydrophilic)





# Choice of MNMs

## Aluminium

2 nanosized forms:

$\text{Al}^0$



Similar size to  $\text{TiO}_2$   
around 20 nm

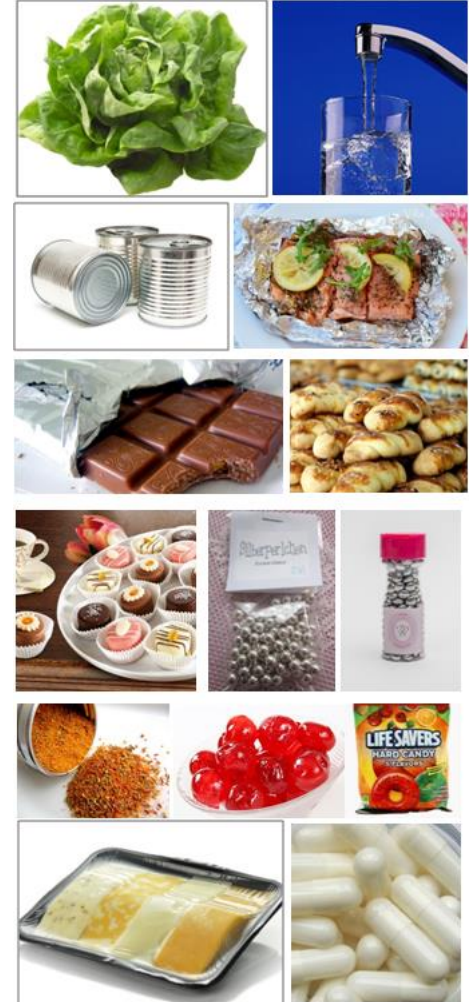
Oxide:  $\gamma\text{Al}_2\text{O}_3$

Comparison with the ionic release by  $\text{AlCl}_3$

# Choice of MNMs

## Aluminium uses

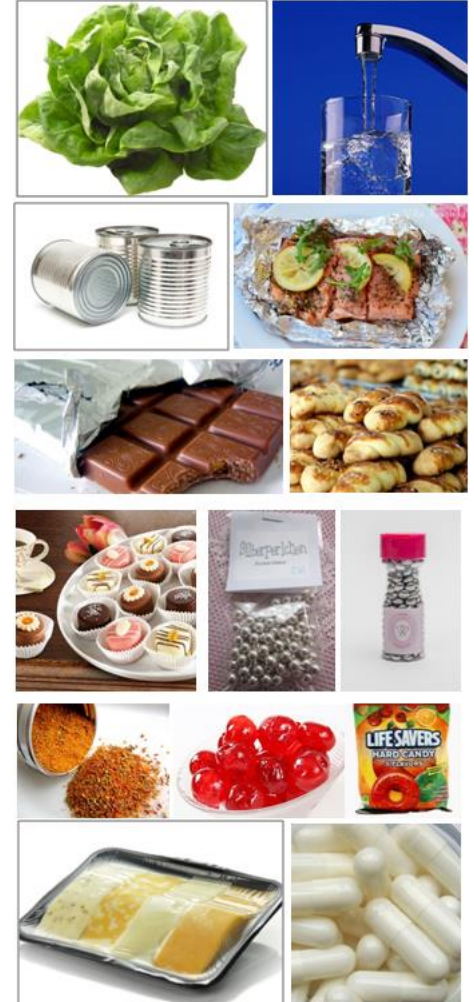
- highly abundant, mostly in oxides, hydroxides or salts
- **natural component** of food and drinking water
- in **direct contact** with food via packages, foils and kitchen ware
- Aluminium-containing **food additives** in many products (confectionary, spices, cheese, candied cherries, biscuits, medical capsules)



# Choice of MNMs

## Aluminium toxicity

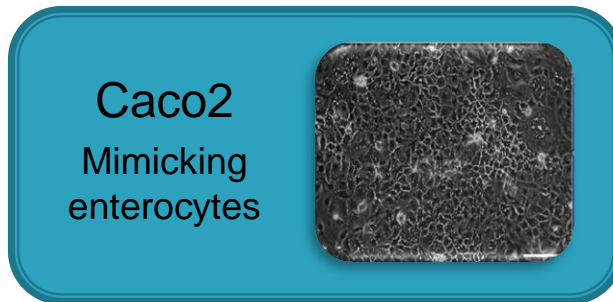
- considered to cause bone diseases, anemia, cancer and neurodegenerative disorders
- uptake, biochemical effects and health hazards are widely unknown



# Choice of models

*In vitro:*

- Cell models of human origin
- Close characteristics to primary cells of the selected tissue
- Good knowledge in the labs
- Representative of the organ of entry (intestine) and the organ of accumulation (liver)

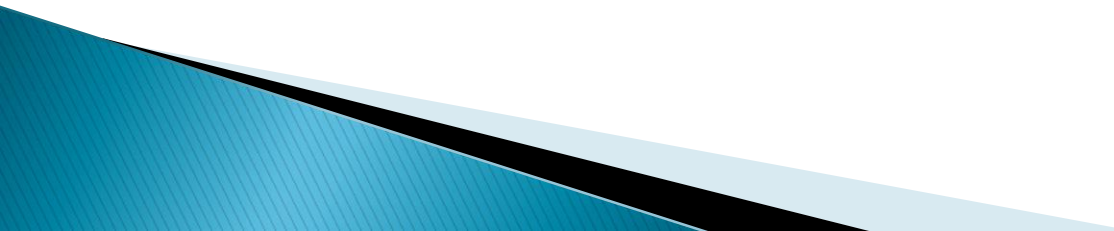


*In vivo:*

Small intestine and liver of rats



# Key issues for the SolNanoTox project

- # Quantification of internal doses of nanomaterials (NM) to estimate Dose-Response relationships
  - # Influence of the human processes like digestion and intestinal mucus secretion on the kinetics, the dynamic behaviour and the biological effects of MNMs?
  - # Stress responses and pathways associated to MNM exposure in liver and intestine?
  - # How solubility impacts NM behaviour and toxicity
  - # *In vivo* extrapolation from *in vitro* biological effects?
- 

# For achievement

Use of **complementary methods** to:

- Measure the **physic-chem characteristics** of NMs
- Detect and quantify the **uptake** into cells and tissues
- Determine the **biological effects** at the molecular, cellular and organ levels with an overview of the cytotoxic and genotoxic responses induced and identification of biomarkers and mechanisms of toxicity

Use of **newly developed and highly performant techniques**

Combination of **integrative *in vitro* and *in vivo* approaches**



Anses

## WP1 Coordination

WP2

BfR

MNMs characterization

Stock solutions  
*ISCR, Mric, ULEI*

Interaction with lipids,  
proteins, cell media,  
intestinal mucus  
*ISCR*

Cellular and tissue internal  
doses and distribution

*BfR, Mric, Anses ULEI*

*In vitro* kinetics  
and digestion

*BfR,*

**WP2**

**Task 2.1  
& Task 2.2**

**Characterization of MNMs in stock solution**

**Interaction of MNMs with cell media and intestinal mucus**

Dispersion protocol of the MNMs:

- **NANOGEN•TOX** protocol (also use in NanoReg)
- in distilled water + 0.05% BSA, sonication during 16 min
- solution of 2.56 mg/ml



Various methods (DLS, NTA, XRD, zeta potential, TEM, Tof SIMS,...) for physic-chemical characterization



## WP2

### Task 2.1 & Task 2.2

## Characterization of MNMs in stock solution

### Interaction of NMNs with cell media and intestinal mucus

**TiO<sub>2</sub>** In stock solution (dispersion medium), data from the European Joint Action Nanogenotox (2010-2013) and the FP7 project NanoReg (2013-2017)

**Al** Behaviour and characteristics in stock solution (dispersion medium) to be obtained

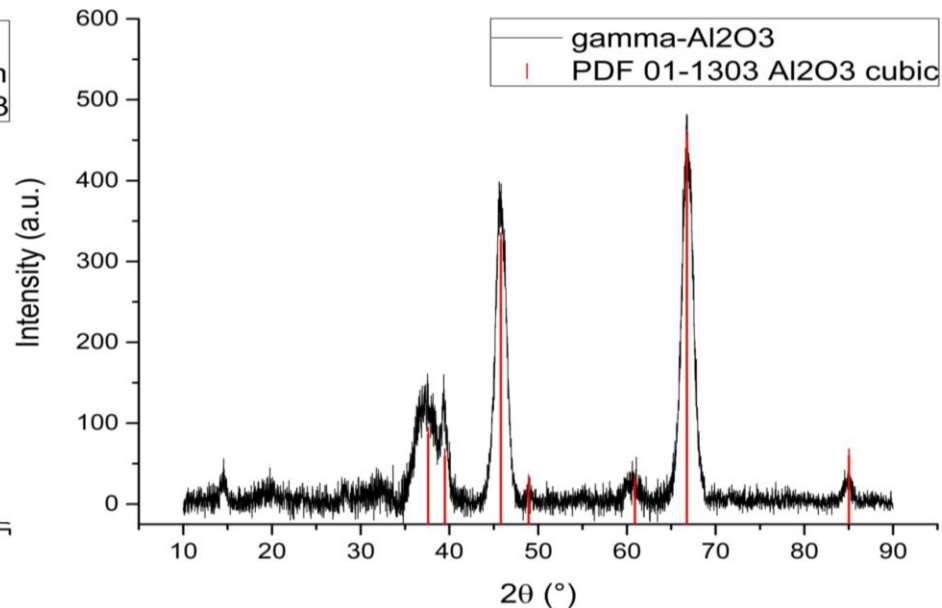
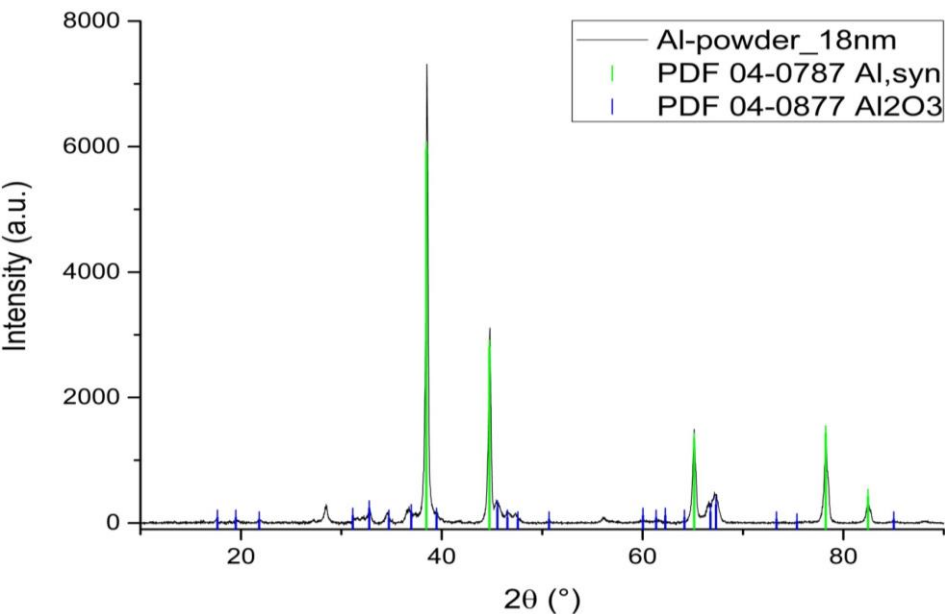
Complete or obtain the data in the cell media used in *in vitro* studies including the stability over time of exposure

Study the interaction with mucus produced by human intestinal cell models *in vitro* and with purified mucins

WP2

Task 2.1  
& Task 2.2

# Characterization of Al NMs

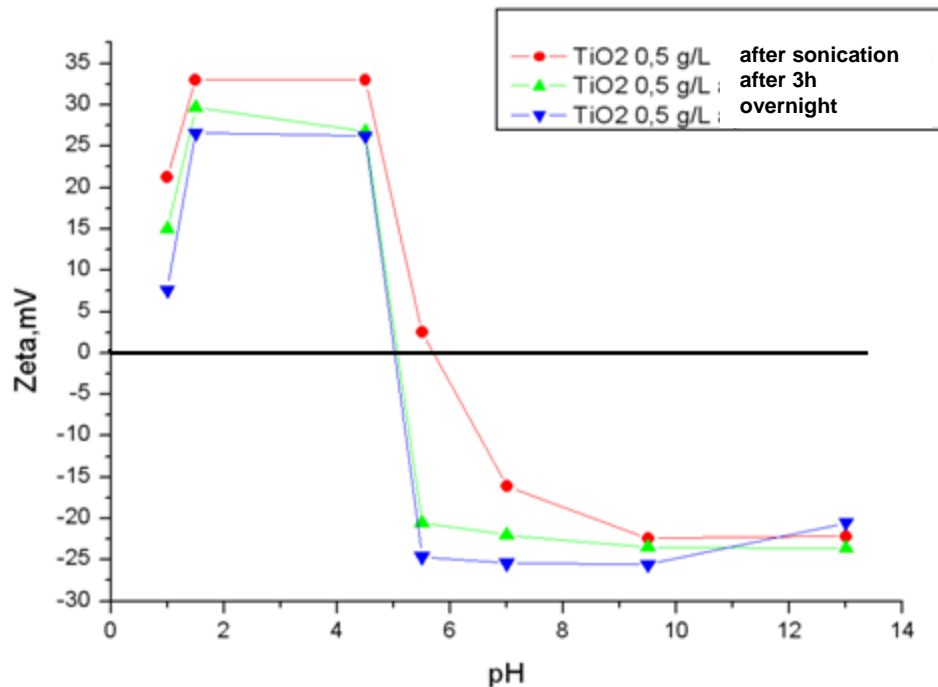


Investigation by X-ray diffraction revealed a thin Al oxide layer at the surface of the metallic Al particles

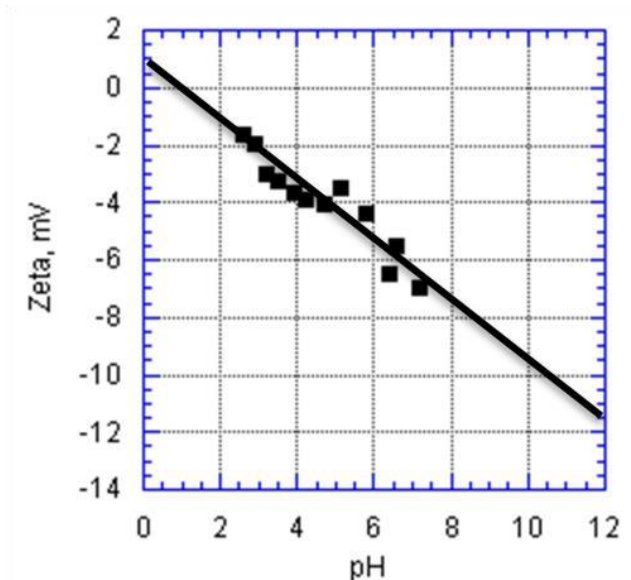
## WP2

### Task 2.1 & Task 2.2

# Interaction of TiO<sub>2</sub> with mucins



Zeta potential of TiO<sub>2</sub> NM-104 with time CNPs=0.5 g/L



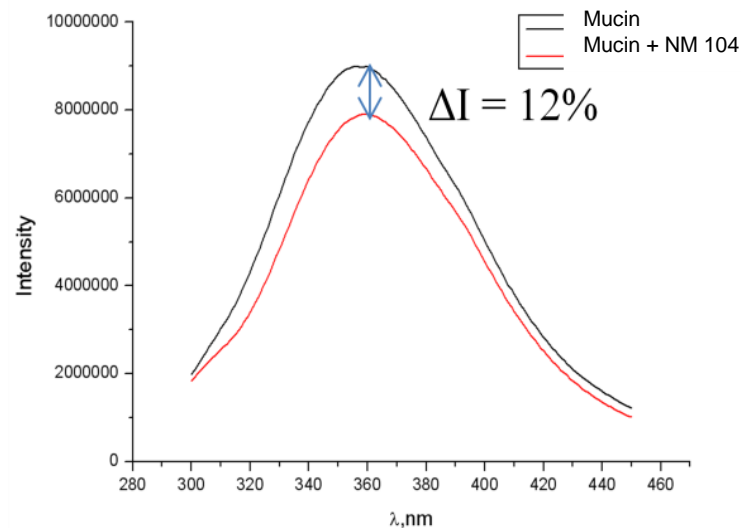
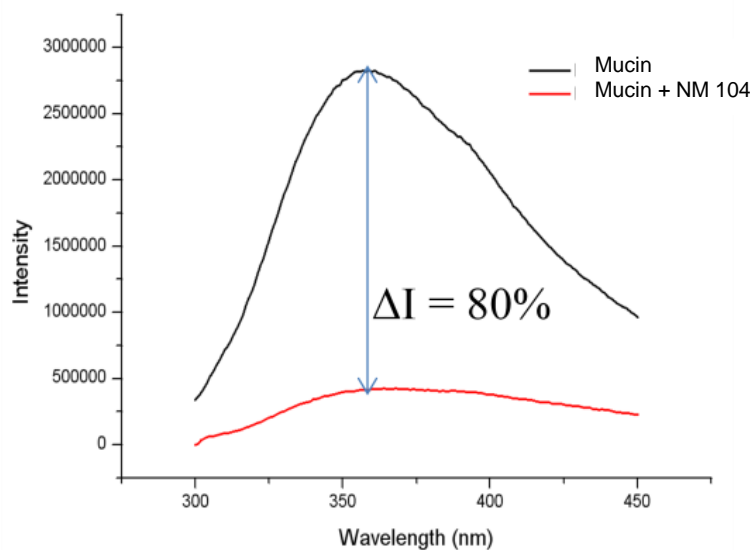
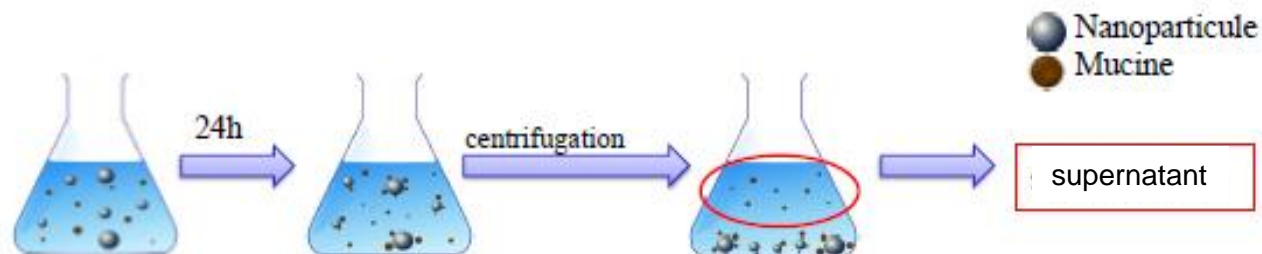
Zeta potential of mucin type III within a pH range (C<sub>mucin</sub> = 0.5 g/L)  
Mucin from stomach of pig

NM104 are positively charged with stomach pH conditions (below 4) and negatively charged in intestine (pH above 6)  
Mucin III is negatively charged independently of pH value

WP2

Task 2.1  
& Task 2.2

# Interaction of TiO<sub>2</sub> with mucins



Fluorescence spectra of supernatants from mucin alone or in the presence of TiO<sub>2</sub> NM-104 at pH 3.5 (left) and pH 8 (right).  $\lambda_{ex}=280\text{nm}$

At pH=3.5 NM104 positively charged adsorbed on mucin which are negatively charged.  
At pH 8 NM104 negatively charged and no interaction with mucin due to repulsion forces.

## WP2

### Task 2.3 & Task 2.4

Internal exposure *in vivo*: Quantification and characterisation of particle uptake in gut and liver

Internal exposure *in vitro*: Quantification and characterisation of particle uptake in gut and liver cells

Methodology:

- TEM
- IBM
- ToF- SIMS
- SP-ICP-MS
- Raman spectroscopy



Use of complementary approaches

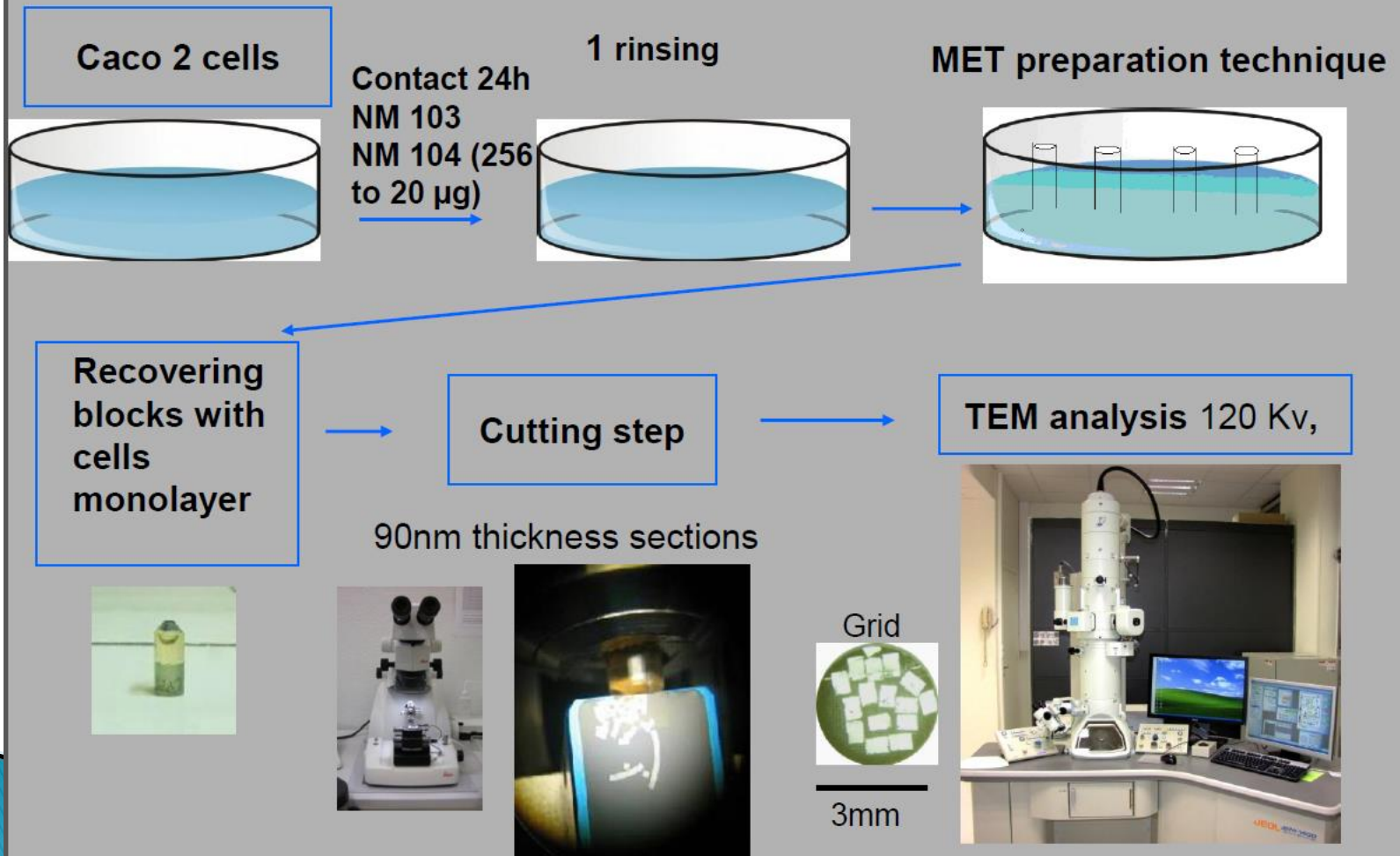
Not only uptake into cells but also distribution inside the cells/ the tissues as well as quantification

WP2

Task 2.3  
& Task 2.4

# Uptake *in vitro*: First results on TiO<sub>2</sub>

## TEM conventionnal approach



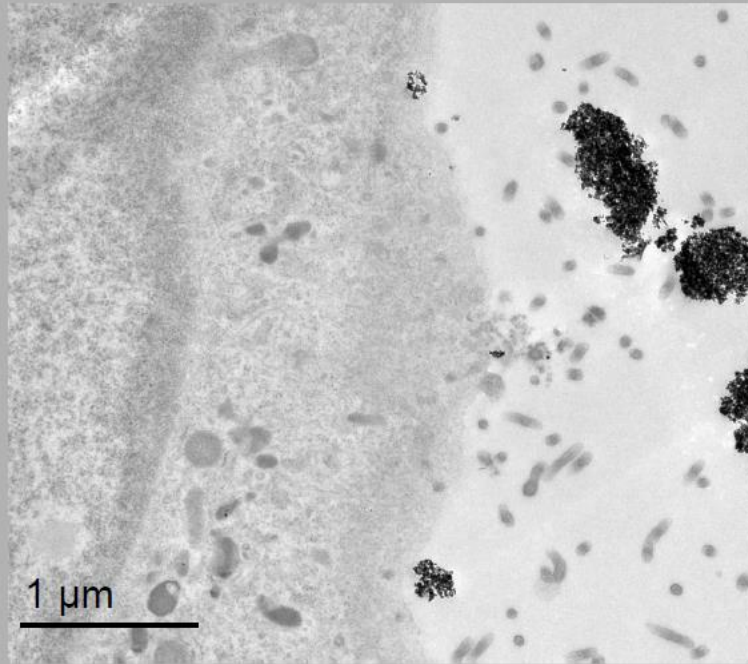
WP2

Task 2.3  
& Task 2.4

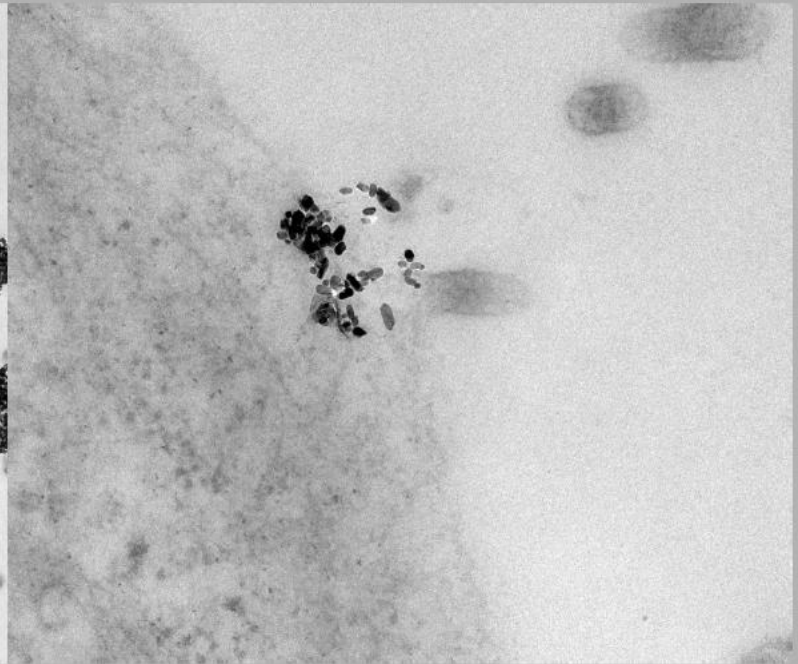
## Uptake *in vitro* in Caco2 cells: First results on TiO<sub>2</sub>

NM entrance into the cells

NM 103-128-15k



NM 103-128-80k



WP2

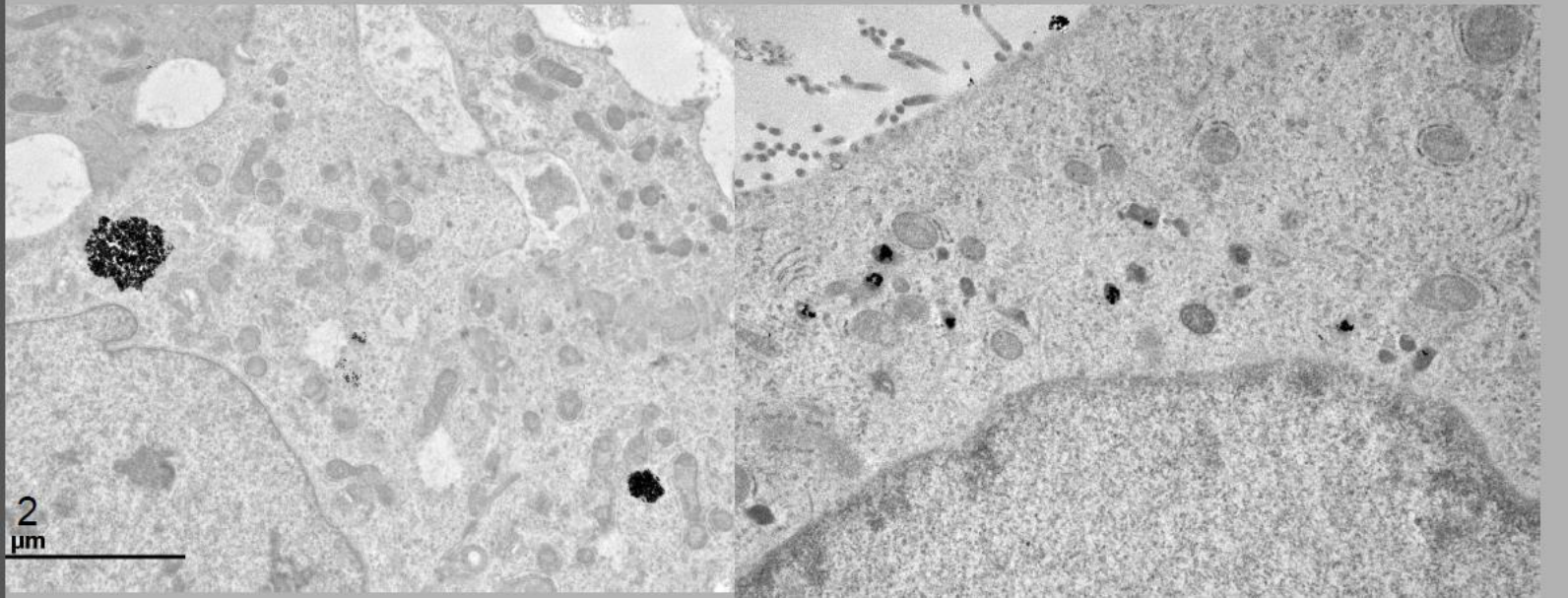
Task 2.3  
& Task 2.4

# Uptake *in vitro* in Caco2 cells: First results on TiO<sub>2</sub>

## NM inside the cells

NM 103-128-8k

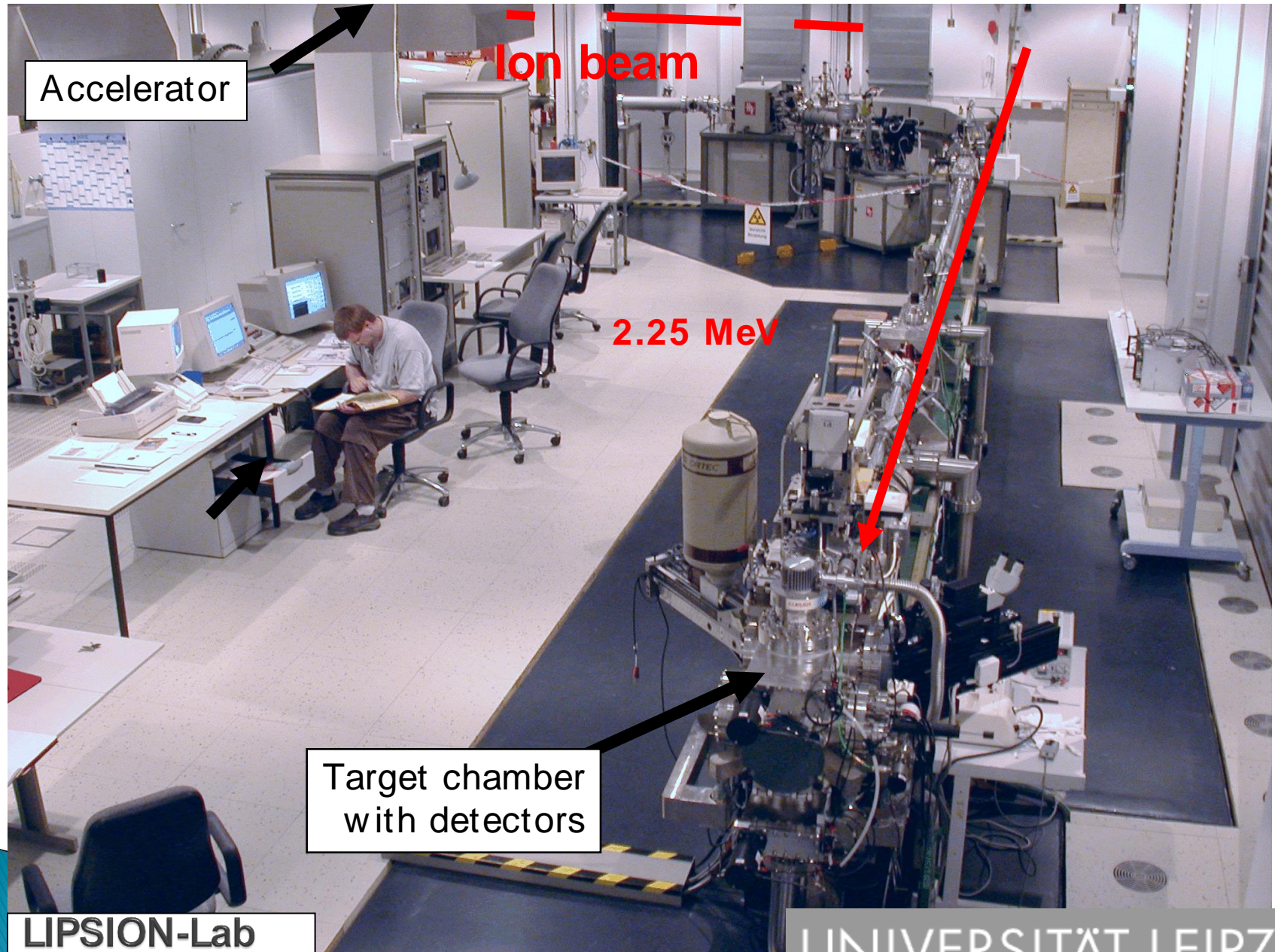
NM 104-256-20k



Both NM103 and 104 whether in free form or inside vesicles (endosomes?)



# The quantification of the NPs uptake *in vitro* and *in vivo* at single cell level by Ion Beam Microscopy

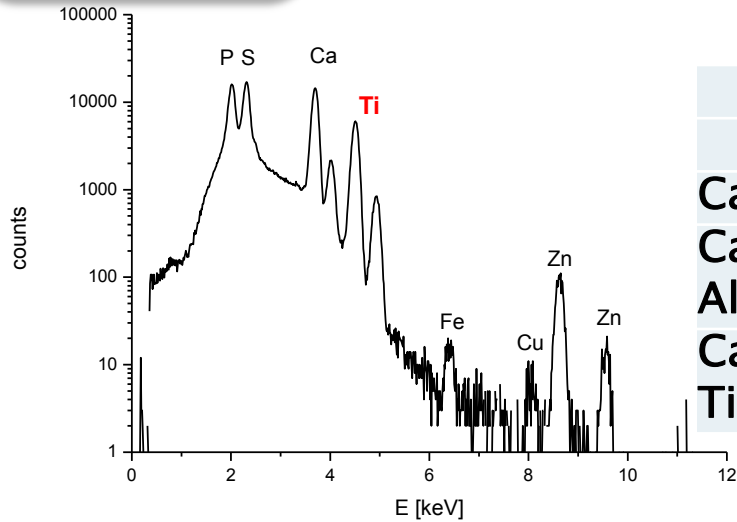


WP2

Task 2.3  
& Task 2.4

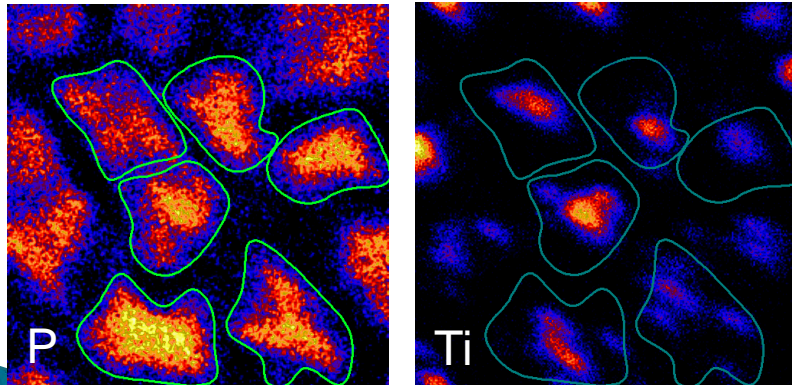
# The quantification of NP uptake in single cells (IBM)

PIXE

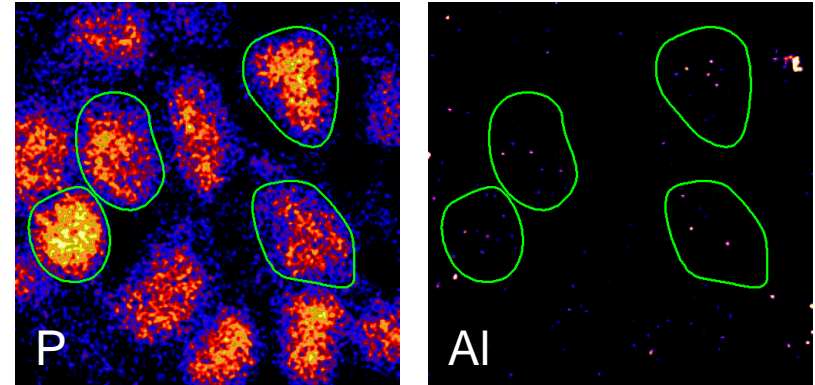


	Elemental Concentration [ppm]		
	P	Ti	Al
Caco-2 Control	13120	0	0
Caco-2 + 50µg/ml Al <sub>2</sub> O <sub>3</sub>	9810	0	230
Caco-2 + 50µg/ml TiO <sub>2</sub>	11500	4790	0

Caco-2 + 50 µg/ml TiO<sub>2</sub> – 24 h exposure



Caco-2 + 50 µg/ml Al<sub>2</sub>O<sub>3</sub> – 24 h exposure

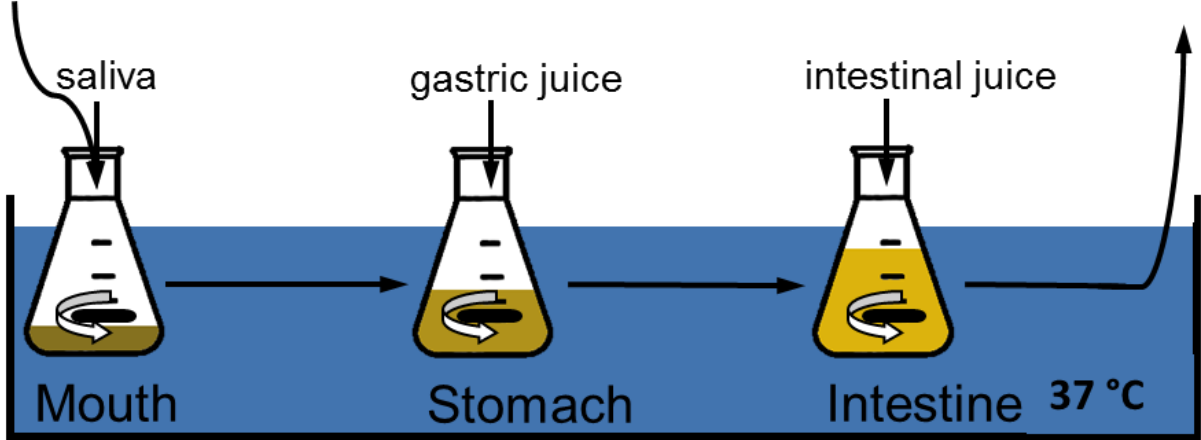


scanfield 50 x 50 µm

**WP2**  
**Task 2.5 &**  
**Task 2.6**

*In Vitro* simulation of different digestic models  
Impact of the digestion process: *In Vitro* Digestion

- NMs can be *in vitro* digested in **artificial fluids**
- simulate way through **gastrointestinal tract**

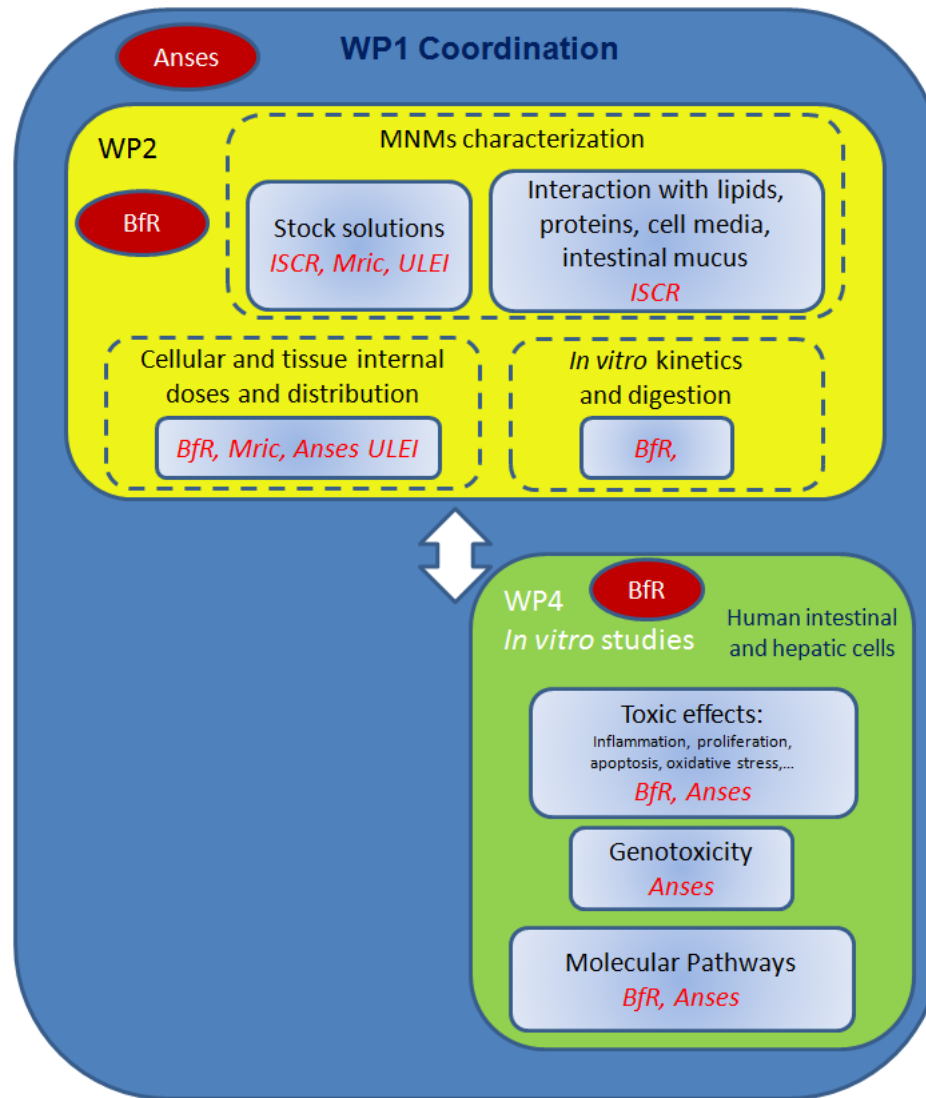


**Characterization  
of digested NMs**

**Uptake and  
transport across  
*in vitro* intestinal  
barrier models**

**Toxicity markers**

**Molecular  
pathways using  
microarrays**



**WP4**  
**Task 4.1**

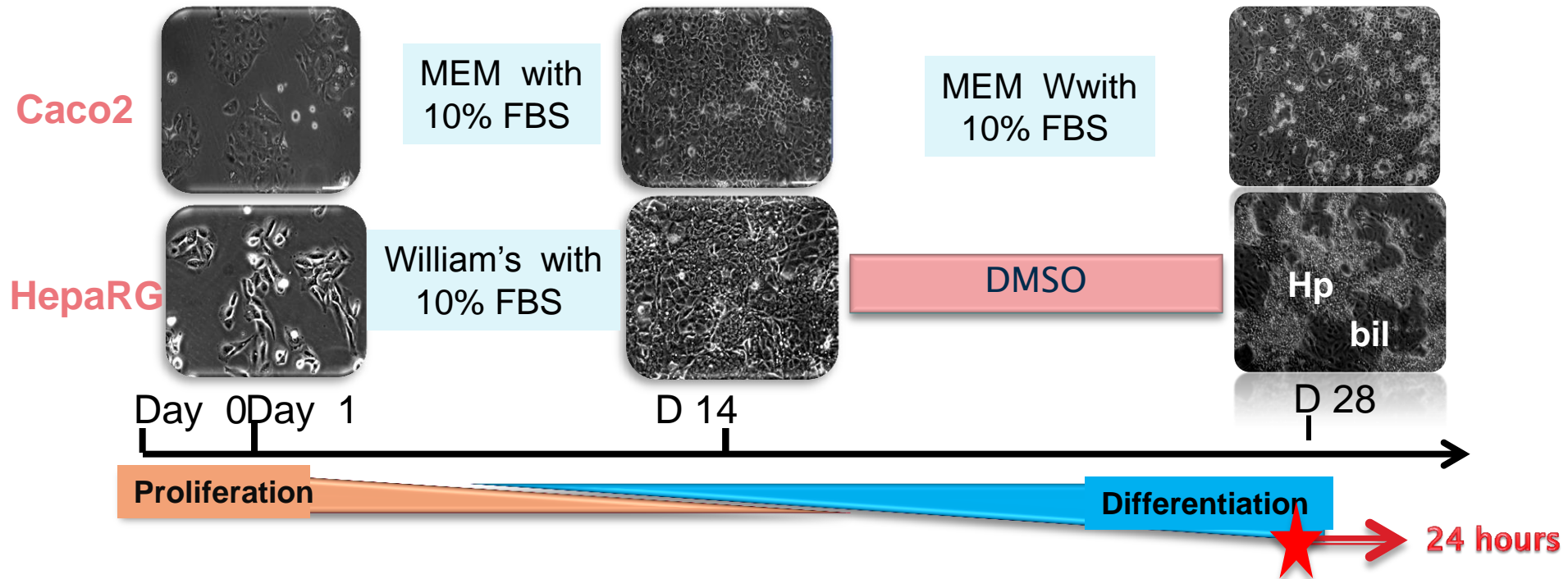
# Cellular response of intestinal and liver cells

- ❖ **Viability**
- ❖ **Morphology**
- ❖ **Other markers (apoptosis, inflammation, oxidative stress,...)**

Newly developed predictive screening methods like High Content Screening and cell impedancy

**WP4**  
**Task 4.1**

# Cellular response of intestinal and liver cells

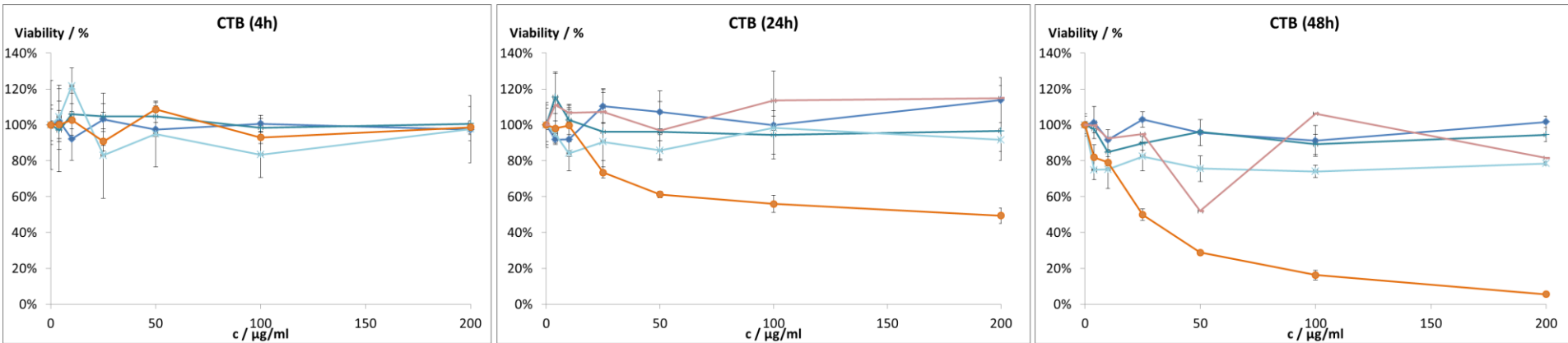


★ **Treatment**  
 NM 103 /NM 104  
 Al  
 256 → 9 µg/ml

- Cytotoxicity: NRU, MTS, casp3 staining (HCS)
- Genotoxicity : comet, micronucleus assay, (γ-H2Ax staining (HCS)
- Cell uptake :TEM, RAMAN, TOF-SIMS...

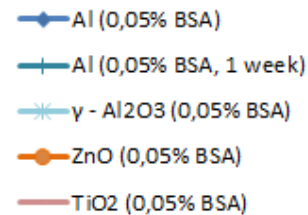
# WP4 Task 4.1

## Cytotoxicity of Nanoparticles on differentiated Caco-2 Cells



### Results:

- Al- and Ti-containing nanoparticles showed apparently no acute cytotoxicity
- Only ZnO particles showed dose- and time-dependent cytotoxicity
- Ionic control substances showed a comparable behaviour



**WP4**  
**Task 4.1**

# Toxic effects: First results on TiO<sub>2</sub>

On differentiated HepaRG

Use of High content analysis to screen several endpoints (at least 3 simultaneously)

Cell count (DAPI), apoptosis (Caspase 3), genotoxicity (H2Ax), inflammation (NFkB), oxidative stress (Nrf2),...



Cells treated for 24h

Fixation and  
Fluorescent  
immunostaining

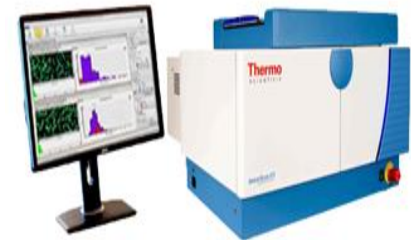
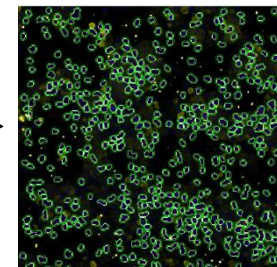
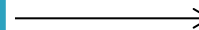
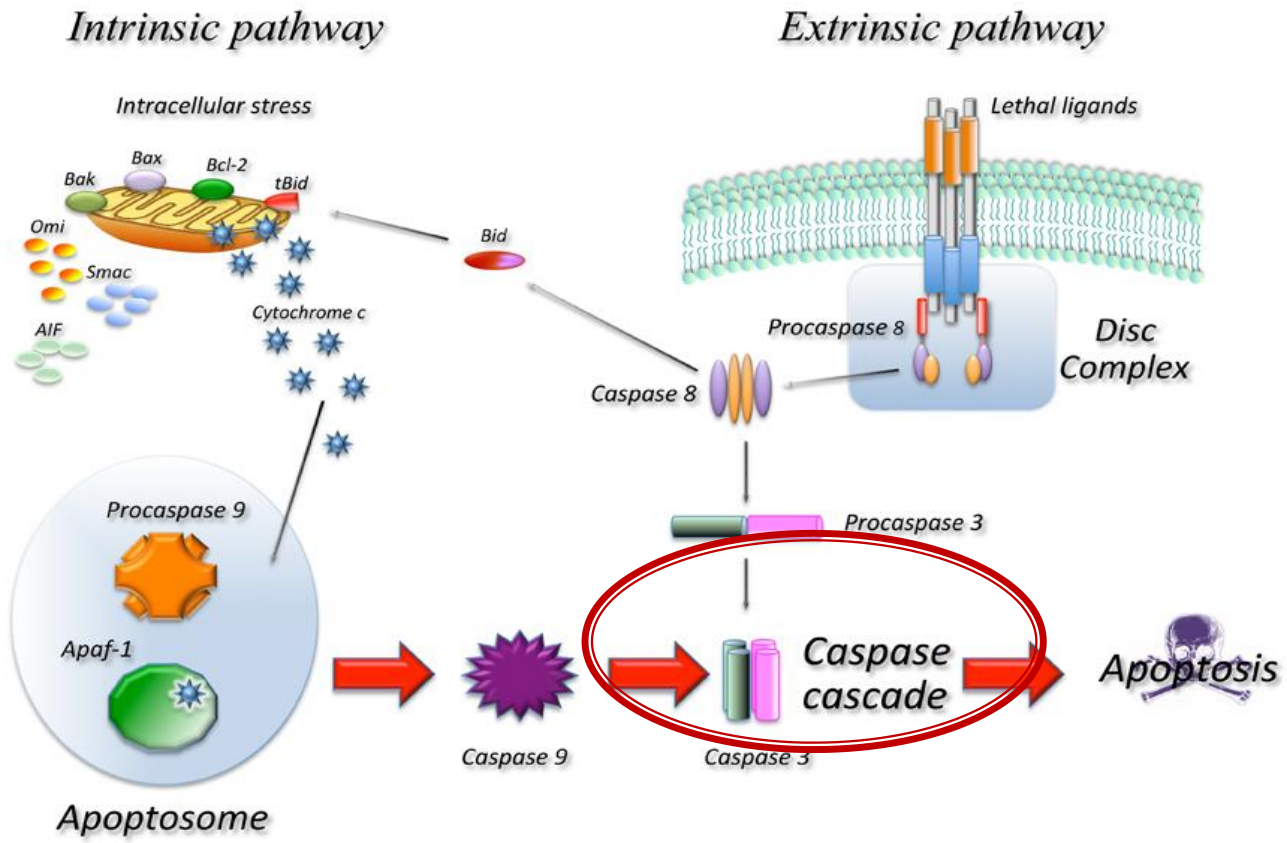


Image analysis



# Toxic effects: First results on TiO2

## Active caspase 3 staining



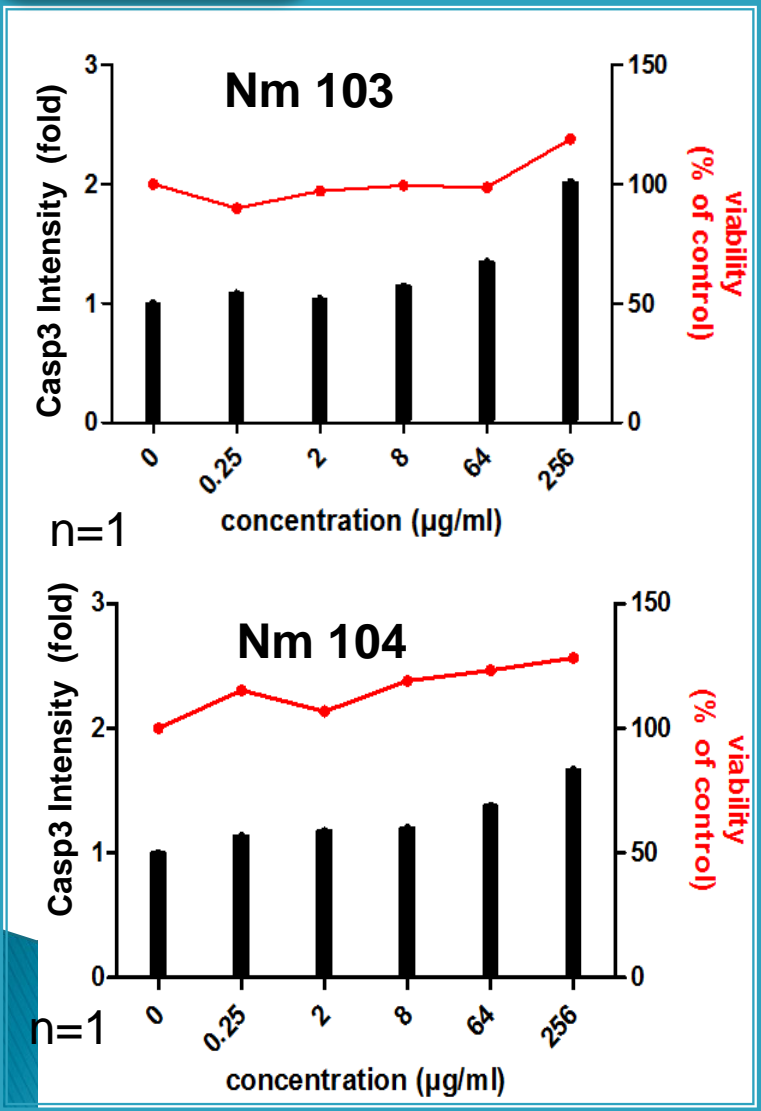
Toxicity → increase of casp3 intensity

**WP4**  
**Task 4.1**

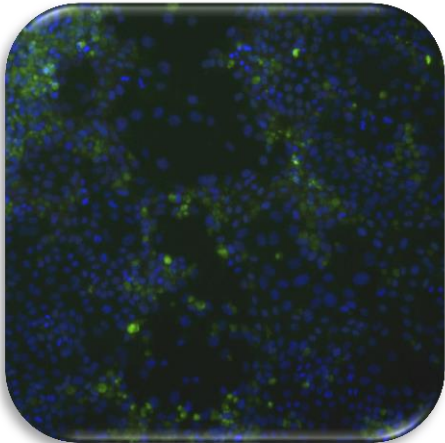
# Cellular response of intestinal and liver cells

## First results on TiO2

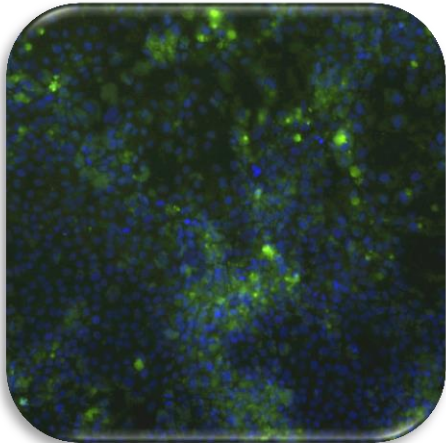
Induction of caspase 3 (apoptosis):  
higher increase induced by NM103



Control



NM103  
256 µg/ml



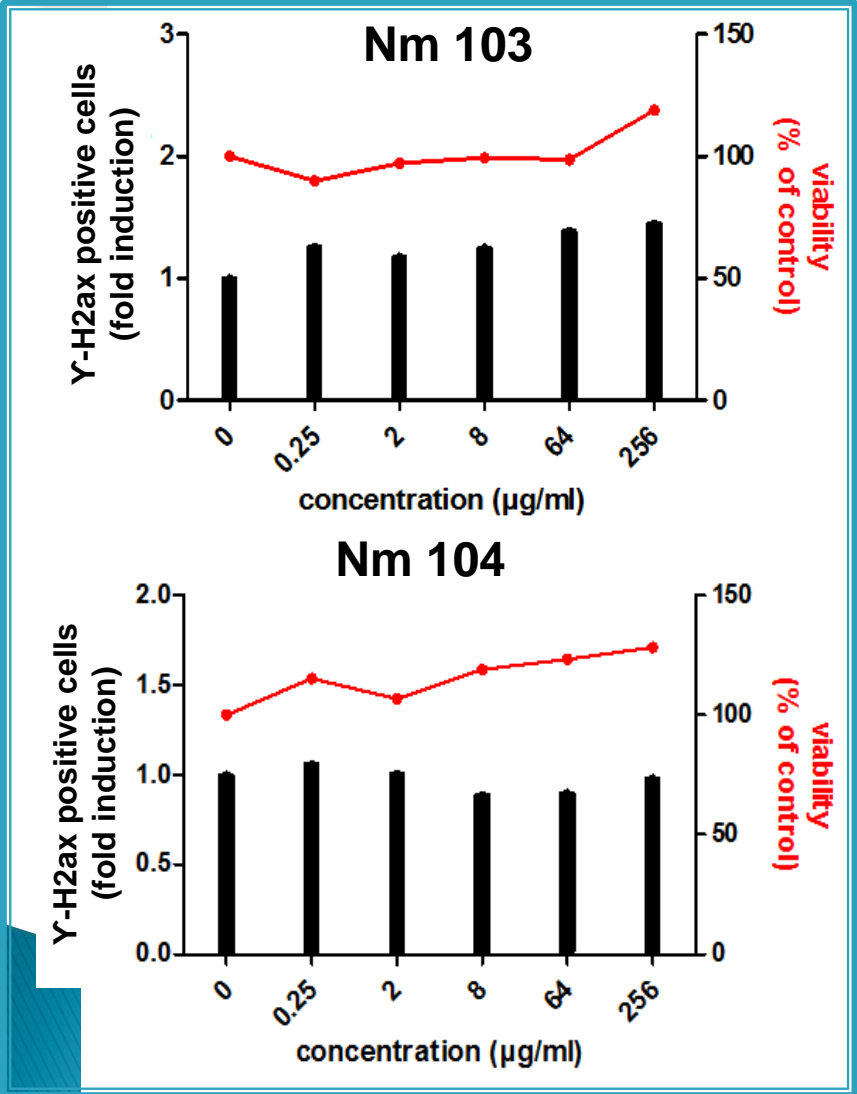
**WP4**  
**Task 4.1**

# Cellular response of intestinal and liver cells

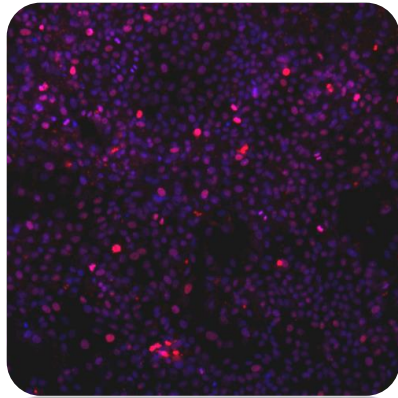
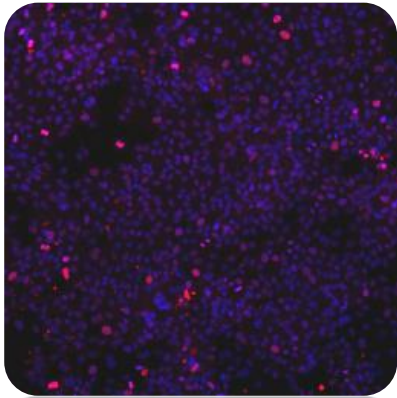
## First results on TiO2

**H2Ax phosphorylated → Double strand break**

No induction of DNA strand breaks with NM103 and NM104



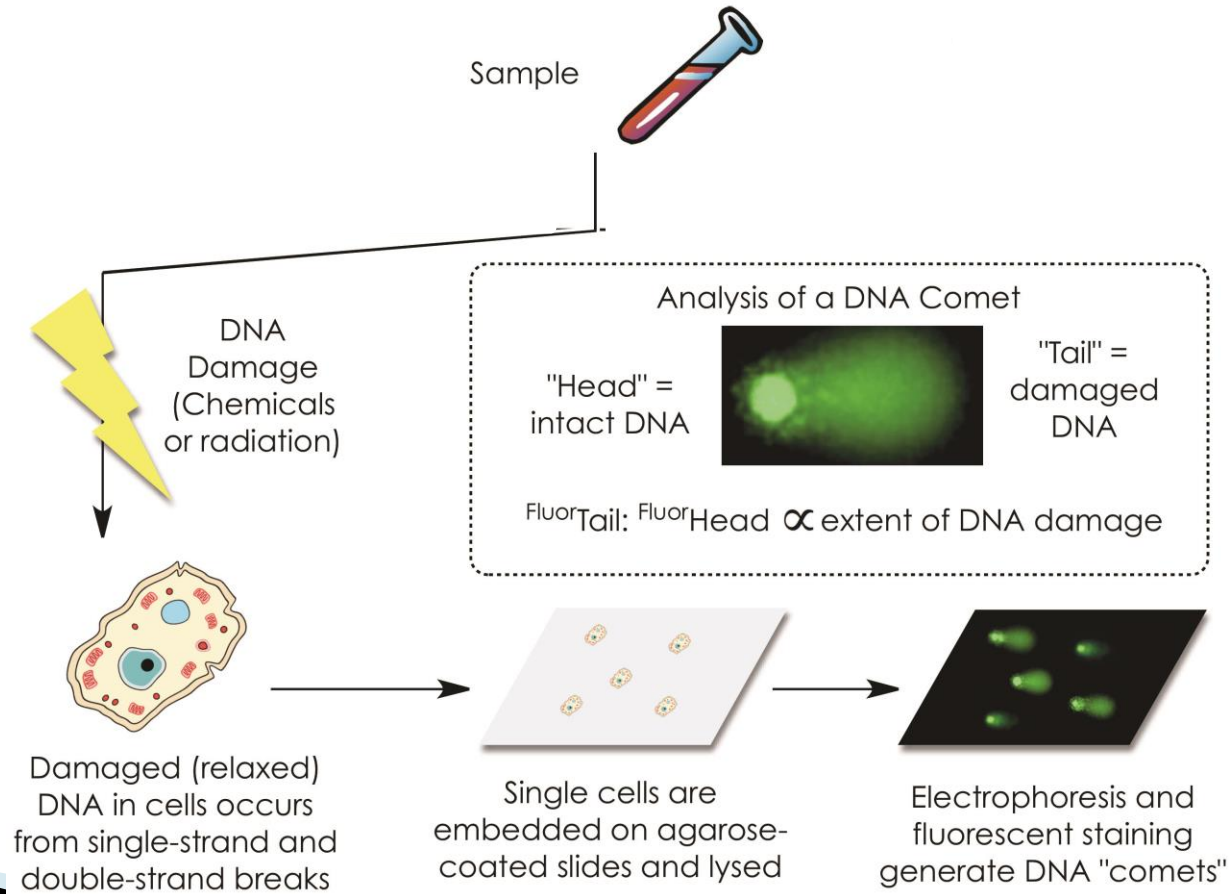
*Control*                      *NM103*  
*256 µg/ml*



**WP4**  
**Task 4.2**

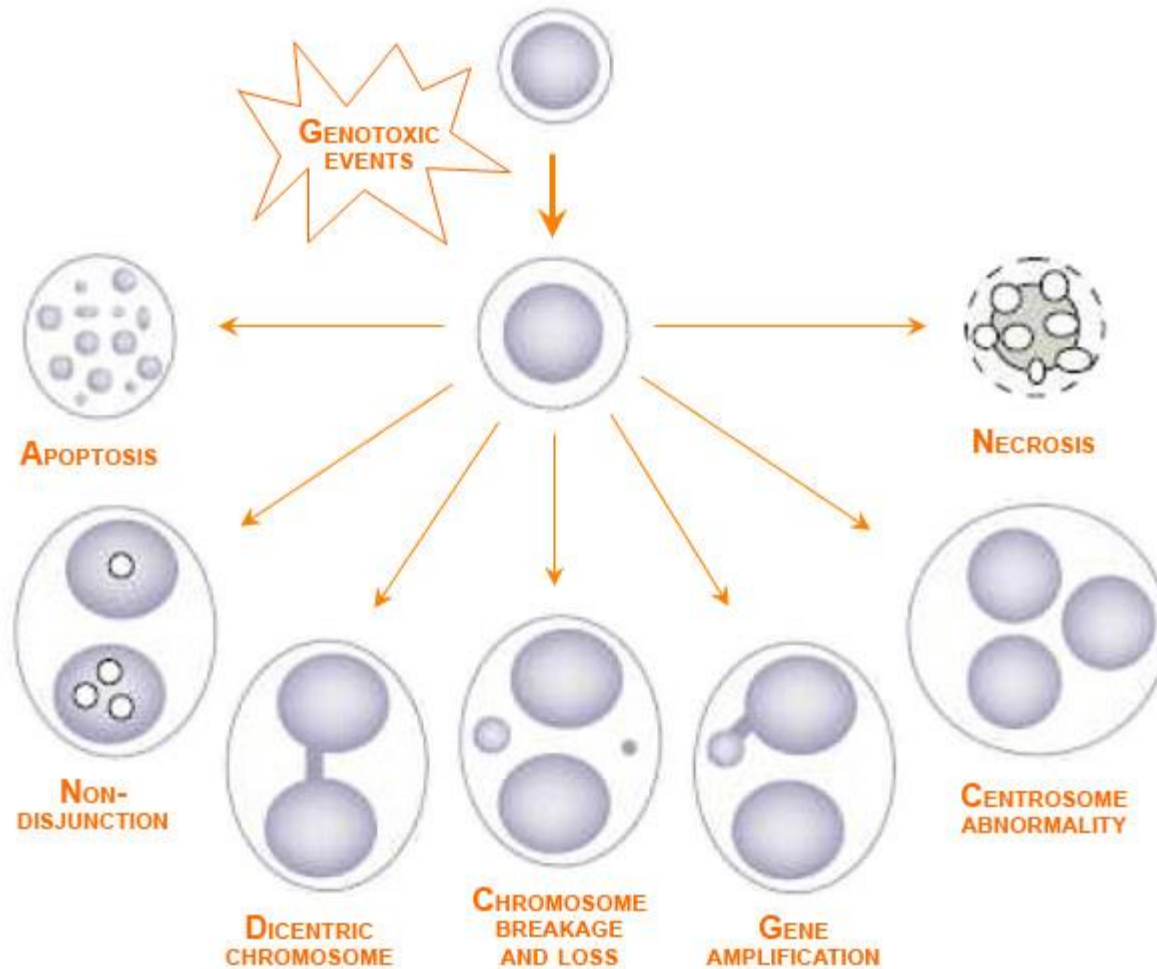
# Genotoxic effects on intestinal and liver cells

## Comet Assay Overview



# Genotoxic effects on intestinal and liver cells

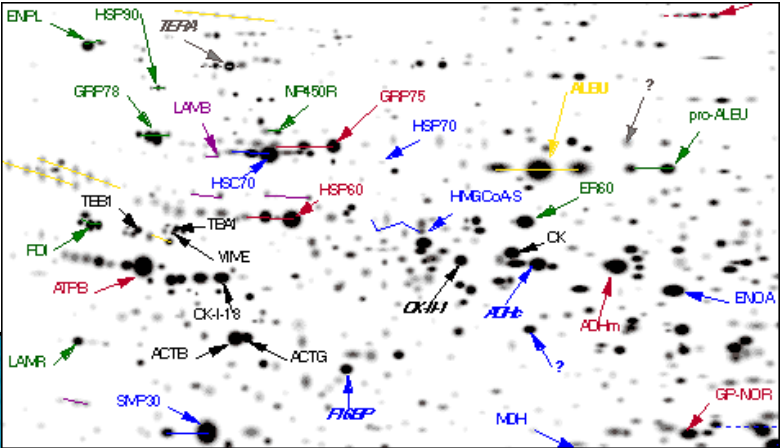
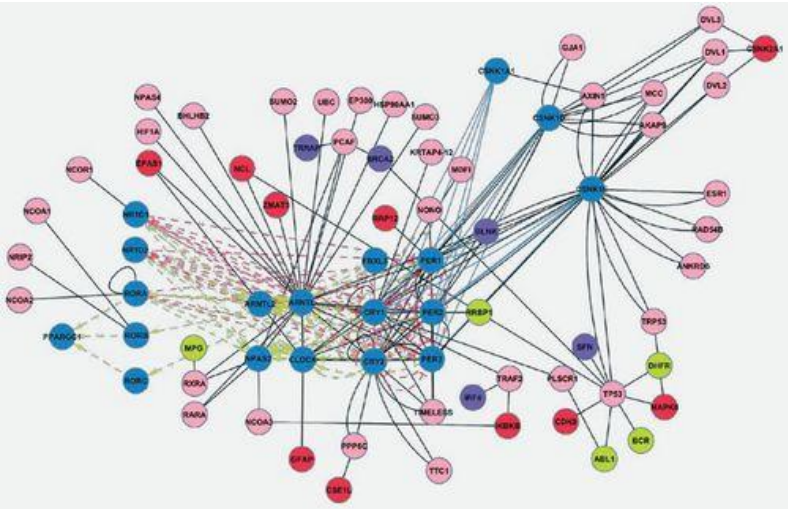
## Cytokinesis-blocked micronucleus assay



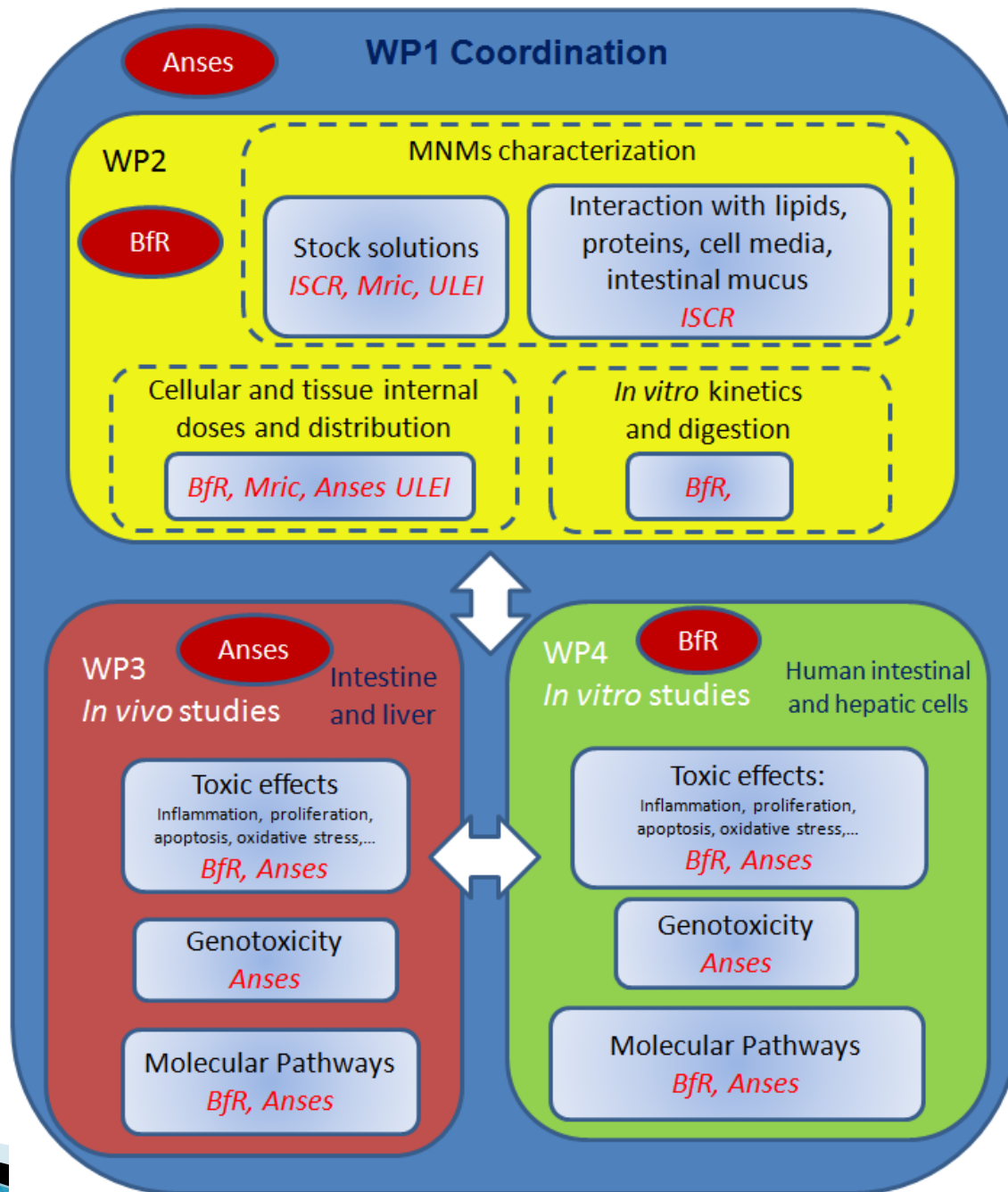
**WP4**  
**Task 4.3**

**Molecular pathways of MNMs toxicity *in vitro***

**Transcriptomics**



**Proteomics**

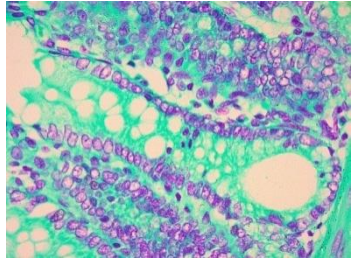
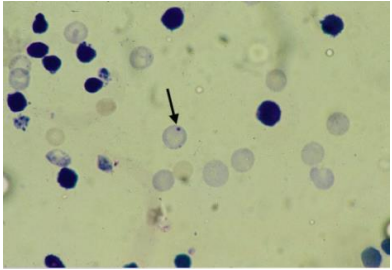
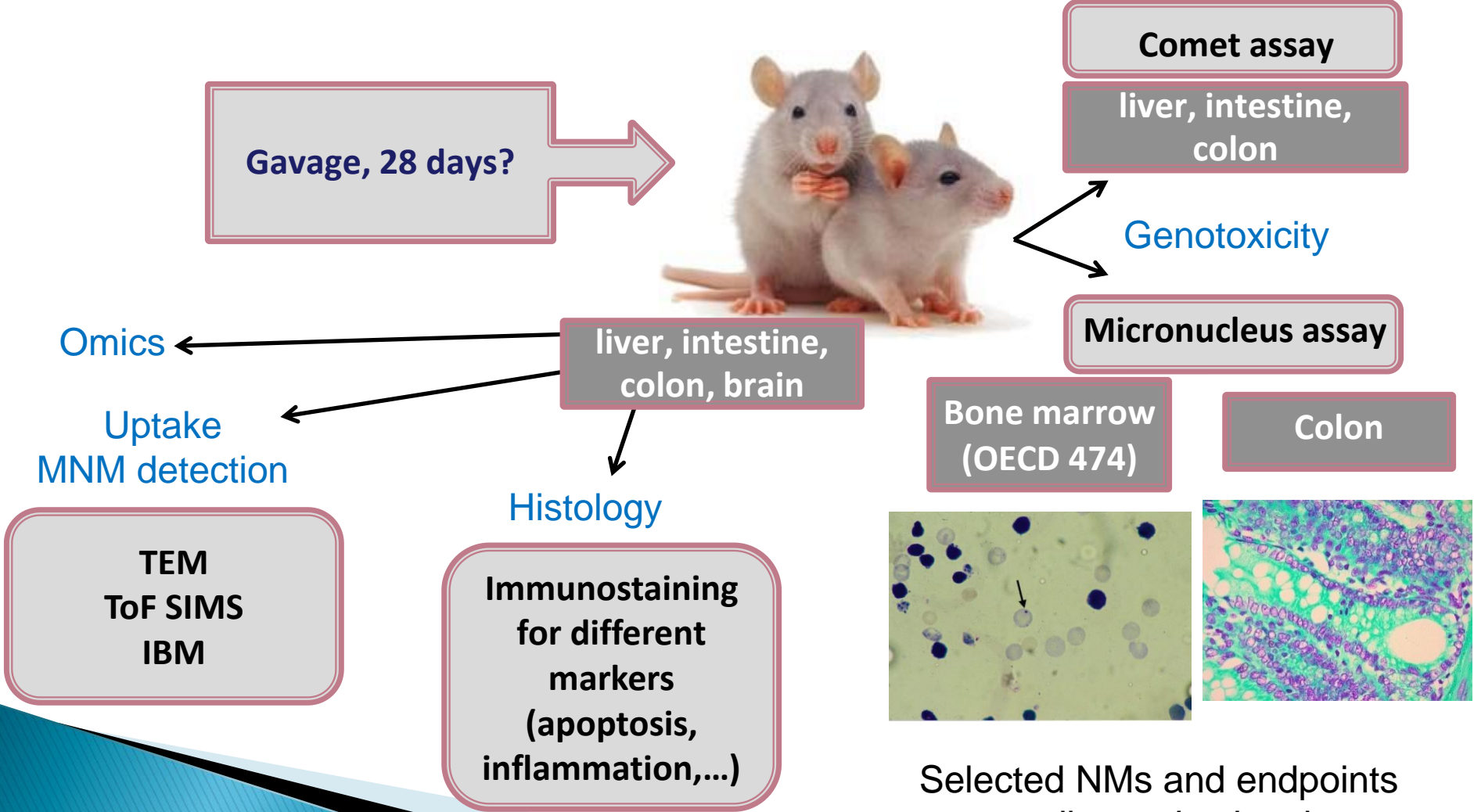


**WP3**  
**Tasks 3.1,**  
**3.2 & 3.3**

# Genotoxicity of MNMs *in vivo*

## Toxic effects of MNMs *in vivo*

### Molecular pathways of MNMs toxicity *in vivo*



Selected NMs and endpoints according to *in vitro* data





**Thank to all the SolNanoTox partners**

Especially the **5 students** (4 PhD and 1 master) :

- Thomas Meyer in ULEI
- Benjamin Krause in BfR
- Hoelger Sieg in BfR
- Pégah Jalili in Anses
- Viktoria Ihnatova in ISCR

# Choice of MNMs

Table 1: Used materials with analytical data

	Al	$\gamma\text{-Al}_2\text{O}_3$	TiO <sub>2</sub> NM-103 Rutil (hydrophobic)	TiO <sub>2</sub> NM-104 Rutil (hydrophilic)	ZnO	AlCl <sub>3</sub> (H <sub>2</sub> O)	ZnCl <sub>2</sub>
Purity	99.9 %	99+ %			99.5 %	99.5 %	99.99 %
APS	18 nm	20 nm	25 nm	25 nm	20 nm	-	-
SSA	40-60 m <sup>2</sup> /g	<200 m <sup>2</sup> /g	51 m <sup>2</sup> /g	56 m <sup>2</sup> /g	50 m <sup>2</sup> /g	-	-
PM	spherical	spherical	-	-	nearly spherical	-	-